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REVIEW

Alzheimer's disease from a diabetic brain: Exploring the molecular process to determine the potential therapy target from marine sources

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Abstract

Background: The prevalence of Alzheimer's Disease (AD) among Type 2 Diabetes Mellitus (T2D) patients has reached almost 30%. Despite the rapid development, treatments for patients with T2D have not succeeded in controlling the neurodegenerative processes that occur in these patients' brains, leading to AD pathology. Several studies have demonstrated that marine sources can inhibit several AD pathogenesis pathways from T2D patients. This review aimed to determine the effect of marine species-sourced compounds at the molecular level in inhibiting and preventing the pathogenesis of AD in T2D patients. Method: Combinations of several terms were used to search for peer-reviewed literature published in PubMed, Scopus, and Google Result: Marine organisms-sourced compounds inhibited various target signaling pathways between T2D and AD, such as an inhibitor of β-site-amyloid precursor protein cleaving enzyme 1 (BACE1) also known as β-secretase, glycogen synthase kinase-3 beta (GSK3B), insulin degradation enzyme (IDE), the mammalian target of rapamycin (mTOR), nuclear factor kappa-light-chain-enhancer (NF-кb), and nuclear factor erythroid 2-related factor 2 (Nrf2). Conclusion: Marine sources can be considered as a potential therapy to prevent or slow AD progression in T2D patients.

Introduction

A major risk factor for cognitive impairment is type 2 diabetes (T2D). Alzheimer's disease (AD) is classified as severe dementia, and this type of cognitive impairment is most common among T2D patients (Yu *et al.*, 2020). There are about 6.5 million elderly individuals suffering from AD. This number is projected to double by 2060 (Alzheimer's Association, 2022). A recent study, that has attempted to discover AD's pathophysiology, revealed that AD is closely related to T2D (Matos *et al.*, 2017). Insulin resistance, neuroinflammation, and oxidative stress are found in the brains of T2D patients (Nguyen *et al.*, 2020). Therefore, T2D is considered the basic mechanism that causes the accumulation of amyloid β (A β) plaque and neurofibrillary tangles (NFT) in AD (Nguyen *et al.*, 2020).

Several studies have demonstrated that marine sources possess compounds that can inhibit several AD pathogenesis pathways from T2D (Li *et al.*, 2013; Gogineni & Hamann, 2018; Ahmad *et al.*, 2019; Lee & Jun, 2019; Arya *et al.*, 2020; Guo *et al.*, 2021). Each of the compounds discovered from marine sources, such as sponges, fungi, algae, tunicates, and coral, has a unique structure and pharmacological activity, and a target to reduce the incidence and severity of AD (Li *et al.*, 2013; Gogineni & Hamann, 2018; Ahmad *et al.*, 2019; Lee & Jun, 2019; Arya *et al.*, 2020; Guo *et al.*, 2021). Therefore, this review aimed to determine the effect of marine compounds at the molecular level in inhibiting and preventing the pathogenesis of AD in T2D.

Methods

Combinations of the search terms "Type 2 Diabetes", "T2D", "Diabetes Mellitus", "Alzheimer's Disease", "AD", "marine source", "pathogenesis", "therapy" and "mechanism" were used to search for peer-reviewed literature published between 2001 and 2022 in PubMed, Scopus, and google scholar. The reference lists of the retrieved articles were examined for additional pertinent articles that were not discovered during the initial search. The initial search found 19 articles, which consisted of randomised controlled trials (RCTs), original articles, systematic reviews, and meta-analyses.

Results

Diabetic individuals have a greater risk of getting AD. Elevated plasma insulin levels are present in both disorders T2D and AD. This event shows that insulin relates to increased plasma A β levels, leading to the formation of A β plaque in the brain (Wrighten *et al.*, 2009). The more specific mechanism of action of marine compounds as anti-Alzheimer can be seen in Table I. Figures 1 to 2 depict the molecular pathways of AD pathogenesis in T2D and the action points of marine source substances in inhibiting the progression of AD in T2D.

Table I: Mechanism of action and active compounds of anti-Alzheimer's disease from marine source

Target of therapy	Marine source	Common name	Active compound	Mechanism of action	References
Inhibitor the β-secretase	Portunus flagicus	Flower crab	Heparan sulfate	Attached to the core of protein form HS proteoglycans (HSPGs)	Mycroft-West et al., 2019
	Sardina pilchardus	While leg shrimp	Glycosaminoglycans	structural destabilisation of BACE1	Mycroft-West et al., 2020
	Ecklonia cava	Brown algae	Eckol, dieckol, 8,8'-bieckol	The side target of BACE1 integrated into the hydroxyl group of the three major phlorotannins formed hydroxyl bonds	Lee & Jun, 2019
	Urechis unicinctus	Innkeeper warm	Hecogenin and cholest-4-en-3-one	-	Naushad <i>et al.,</i> 2019
GSK-3 inhibitor	Ircinia dendrides and Callyspngia truncate	Marine sponge	Palinurin and Phenylmethylene	Non-competitive inhibitors at the ATP-binding site.	Arya <i>et al.,</i> 2020
	Ircinia dendrides	Marine sponge	Manzamine A	The presence of a double bond between positions 15 and 16 increased the capability of manzamine to inhibit the GSK-3	Kubota <i>et al.</i> , 2020
	Aplidium meridianum	Tunicate	Meridianin	The hydroxyl group in position 4 and the substitution of Br in position 6 in the indole ring inhibited the GSK-3.	Alonso & Martnez, 2006
	Botryotinia fuckeliana	Marine fungus	Pannorin, Alternariol, and Alternariol monomethylether	Their benzocoumarine ring plays a role in transferring electrons to make GSK-3 inhibition more efficient	Wiese <i>et al.</i> , 2016
Insulin- degradation enzyme (IDE)	Streptomyces tumescens	Marine bacteria	Tumescenamides A and B	Induces the reporter gene expression and controls the IDE	Gogineni & Hamann, 2018
Nuclear- factor erythroid 2- related factor	Erylus formasus	Marine sponge	Triterpenoid glycoside	Induce the antioxidant response element (ARE)	Li, Himaya & Kim, 2013
	Holothuria scabra	Sandfish (sea cucumber)	Triterpenoid glycoside	Induce the antioxidant response element (ARE)	Li, Himaya & Kim, 2013
NF-ĸB Inhibitor	Trididemnum solidum	Tunicate	Didemnin B (Depsipeptides)	LPS-induced macrophages, and inflammatory mediators	Ankisetty <i>et al.</i> , 2013; Ahmad <i>et</i> <i>al.</i> , 2019;; Ghiciud <i>et al.</i> , 2021
	Serratia marcescens, Vibrio psychroerythrus, Pseudoalteromonas denitrificans and Zooshikella rubidus	Marine bacterium	Prodigiosin and Cycloprodigisin	Inhibits TNF-α-induced NF-κB activation	Kamata et al., 2001; Ahmad et al., 2019;

Target of therapy	Marine source	Common name	Active compound	Mechanism of action	References
	Bacillus sp. HC001, Piscicoccus sp. 12L081	Marine bacterium	Diketopiperazines	Downregulation of TNF- α , IL-6, NF- κ B, and ERK1/2	Ahmad <i>et al.</i> , 2019
	Axinella verrucosa, Acanthella aurantica and Stylissa massa	Sponge	Alkaloids	NF-kB-specific inhibitors, inhibit the production of IL-8, IL-2, and TNF- $\!\alpha$	Ahmad <i>et al.</i> , 2019
	Ishige okamurae	Algae	Diphlorethohydroxyc ar malol	Suppress the expressions of p-p65 NF-κB	Kim <i>et al.,</i> 2020
	Ecklonia cava	Algae	Dieckol	Inhibits the increase of inflammatory markers	Barbalace <i>et al.,</i> 2019
	Ecklonia stolonifera	Algae	Phlorofucofuroeckol B	Inhibited the phosphorylation of Akt, ERK, and JNK	Barbalace <i>et al.,</i> 2019
	Gracilaria verrucosa	Algae	(E)-9-Oxooctadec-10- enoic-acid and (E)- 10-Oxooctadec-8- enoic-acid	Suppress the production of inflammatory biomarkers including NO, IL-6, and TNF- $\!\alpha$	Ahmad <i>et al.,</i> 2019
	Aspergillus insulicola	Hawaiian marine sedimentary fungus	Azonazine (hexacyclic dipeptide)	inhibiting the production of NF-κB	Ahmad <i>et al.,</i> 2019
	Penicillium paxilli Ma(G)K	Fungi	Pyrenocine A	-	Ahmad <i>et al.,</i> 2019
mTOR Inhibitor	Fascaplysinopsis sp.	Marine Sponge	4-chloro fascaplysin	Inhibited cell survival through PI3K/Akt/mTOR pathway	Sharma <i>et al.,</i> 2017
	Brown seaweeds	Seaweeds	Fucoxanthin	Inhibitory effect on the Akt/ mTOR signalling	Guo <i>et al.</i> , 2021
	Sinularia flexibilis	Soft Coral	Sinulariolide	Inhibiting the phosphorylation of PI3K, Akt, and mTOR	Li <i>et al.</i> , 2013
	Streptomyces hygroscopicus	Marine bacterium	Sirolimus	Binds to FK-bound protein 12 and serine- threonine kinase, mTOR, inhibiting the transduction of IL-2R	Nguyen <i>et al.,</i> 2019

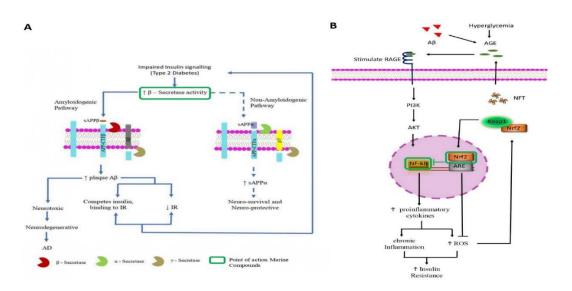


Figure 1: A) The basic mechanism between T2D and AD. Marine compounds act as β-secretase inhibitors. B) Molecular Pathway Effect of T2D on increased oxidative stress and proinflammatory cytokines. Marine compounds act as NFkB inhibitors and Nrf2 inducers. Insulin Receptor (IR); Alzheimer's Disease (AD), Amyloid β (Aβ); soluble Amyloid Precursor Protein by α secretase (sAPPα); soluble Amyloid Precursor Protein by β secretase (sAPPβ); Advanced Glycation End Products (AGE); Receptor for Advanced Glycation Endproducts (RAGE); Nuclear-factor erythroid 2-related factor (NRf-2); Reactive Oxygen Species (ROS); Antioxidant Responsive Element (ARE); Kelch-like ECH associated-protein 1 (Keap1); Neurofibrillary tangles (NFTs)

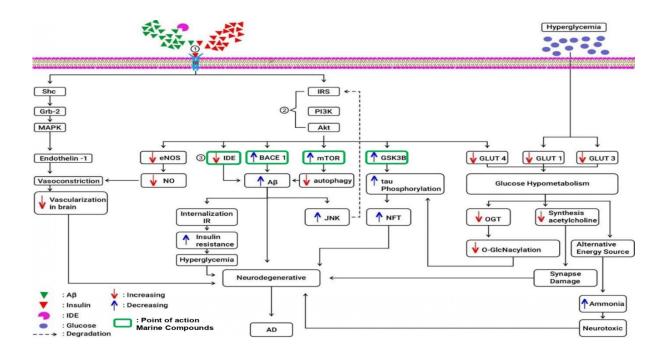


Figure 2: Molecular pathway of impaired Insulin Signaling and glucose metabolism in T2D and AD. Marine species-sourced compounds act as β-secretase inhibitors, mTOR inhibitors, and GSK3B inhibitors, and increase IDE expression. Insulin receptor (IR); insulin receptor substrate (IRS); phosphatidylinositol 3-kinase (PI3K); protein kinase B (Akt); insulin-degrading enzyme (IDE); beta secretase-1 (BACE1); Amyloid β (Aβ); mammalian target of rapamycin (mTOR); glycogen synthase kinase-3 beta (GSK3B); glucose transporter (GLUT); c-Jun N-terminal kinase (JNK); neurofibrillary tangle (NFT); endothelial nitric oxide synthase (eNOS); nitric oxide (NO); src homology and collagen protein (Shc); growth factor receptor bound protein 2 (Grb-2); mitogen-activated protein kinase (MAPK); Alzheimer's disease (AD)

Discussion

Accumulation of A β , a peptide derived from amyloid precursor protein (APP), is one of the hallmarks of Alzheimer's disease (AD). There are two pathways for APP degradation; amyloidogenic and nonamyloidogenic (Figure 1A). Normally, most APP is cleaved by non-amyloidogenic pathways (Najem *et al.*, 2016). Insulin was shown to enhance the nonamyloidogenic pathway by stimulating α -secretion activity. Additionally, sAPP α , a protein resulting from non-amyloidogenic pathways, is involved in neuronal survival and protection (Shieh *et al.*, 2020; Soriano *et al.*, 2001).

In diabetic individuals, impaired insulin signalling results in an increase in β -secretase activity and a decrease in α -secretase activity (Shieh et~al.,~2020). Thus, the APP breakdown pathway becomes more amyloidogenic than non-amyloidogenic. This event will cause a decrease in sAPP α , a neuroprotective protein, and an increase in A β oligomers (Shieh et~al.,~2020). A β oligomers have been shown to impair synaptic plasticity in neurons by inhibiting long-term

potentiation (LTP) and long-term depression (LTD). Additionally, the accumulation of $A\beta$ oligomers facilitates the formation of neurotoxic $A\beta$ plaques, resulting in neuronal death and synaptic injury (Lei *et al.*, 2016; Varga *et al.*, 2015).

Many marine sources were reported to inhibit the β -secretase or disease-relevant β -secretase 1 (BACE1) (Table I). The four species have each active compound with a mechanism action in BACE1. The active compounds were glycoside, flavonoid and steroid groups. Nevertheless, the final action of these compounds was to inhibit the BACE1 activity. It could reduce the accumulation of the A β plaque and the progression of AD.

Aβ oligomers also worsen insulin signalling impairment in T2D by degrading the Insulin Receptor Substrate (IRS), decreasing the number of Insulin receptors (IR) (Figure 2), and competing with insulin to bind to the insulin receptor. Shieh et al. (2020) and Tumminia et al. (2018) showed that disruption of the insulin signalling pathway in T2D impairs the PI3K-Akt cascade's activation. This impaired insulin signalling results in several events that aggravate AD (Figure 2), some of

which are: 1) decreased secretion of insulin-degrading enzyme (IDE); 2) abolishment of the inhibitory effect of Glycogen Synthase Kinase 3 Beta (GSK3B); 3) elimination of insulin's inhibitory action on mammalian target of rapamycin (mTOR). IDE plays a role in A β degradation. Thus, decreased IDE secretion leads to further A β accumulation (Tumminia *et al.*, 2018; Nguyen *et al.*, 2020).

The activation of GSK3B plays a role in tau protein phosphorylation and causes tau protein accumulation and NFT formation. The accumulation of NFT causes impaired synapse formation, neuronal cytoskeletal collapse, and neurite retraction (Tumminia $et\ al.$, 2018; Nguyen $et\ al.$, 2020). Notably, mTOR activation inhibits normal autophagy processes. At the same time, the autophagy process is needed to degrade various abnormal proteins, including A β and tau proteins (Cai $et\ al.$, 2015).

The compounds that were reported to have the activity to inhibit the GSK3B activity and induced IDE activity were found in marine sources. Streptomyces tumescens was the only reported marine source that induced the IDE activity (Table I). Tumescenamides A was the type of peptide found in S. tumescens that can increase the presence of IDE. Tumescenamide A and B can induce the upregulation of the IDE enzyme, a metalloprotease enzyme that is responsible for insulin degradation. It proved to have a role in A β degradation which is expected to be a promising treatment of AD (Motohashi et al., 2010).

Most species of these marine sources were found in the sponge (Table I), tunicate and marine fungi also contain compounds that can inhibit the activities of the GSK3B (Alonso & Martnez, 2006). The compounds that play the role of inhibiting the GSK3B included the sesquiterpene, alkaloid and polyphenolic groups. The mechanism of action of each compound was different, but the final action was to inhibit the GSK3B activity. Palinurin, a sesquiterpene from types of marine sponges exerts its high selectivity inhibition of the allosteric site of GSK3B (Hafez & Kijjoa, 2021).

Four species that were reported can inhibit the mTOR (Table I). Brown seaweeds and sponges were marine sources that were easy to find in the sea. The type of compounds in these marine sources was alkaloid, macrolide, carotenoid, and diterpene lactone. The mechanism of action of these compounds was to inhibit the upstream mTOR pathways. Fucoxanthinol, the active form of fucoxanthin metabolite from seaweeds has a neuroprotective activity by preventing inflammation, ROS scavenging effect, and activation of mTOR (Song & Zhou, 2022). Sirolimus can be found in the marine bacterium *Streptomyces hygroscopicus* (Nguyen *et al.*, 2019).

Stress oxidative and neuroinflammation are other hallmarks that play roles in AD pathogenesis (Figure 1B). Hyperglycemia patient causes the production of advanced glycation end products (AGE) in diabetic patients. AGE is formed when proteins or lipids are glycated with glucose or its metabolites. The binding of AGE with the Receptor for Advanced Glycation End Products (RAGE) will promote the activity of Nuclear Factor kappa B (NF-kB), which will stimulate various proinflammatory cytokines, including IL-β and TNF-α and increase Reactive Oxygen Species (ROS) (Granic et al., 2009). Aβ can also become AGEs and bind to RAGE, aggravating oxidative stress and chronic neuroinflammation (He & Sun, 2021).

The NRf-2 has a relationship with antioxidant activity. The increase of NRF-2 can activate the antioxidant transcription. The seaweeds, marine macroalgae, contained sulfated polysaccharides with potent antioxidant activity (Cardoso *et al.*, 2016). Byun *et al.* (2018) showed that polysaccharides mediated the Nrf-2 activation (Table I). The species of marine source that inhibited the NFkB was algae (Table I). Various compounds were found in algal species, such as polyphenols and fatty acids. The common mechanism of action of these compounds decreased the release of inflammation mediators (Byun *et al.*, 2018).

Conclusion

Marine sources possess potential therapies to prevent or slow AD progression in T2D patients. Marine species-sourced compounds have mechanisms of action that inhibit the progression of Alzheimer's disease in T2D patients. Due to impaired insulin signalling, some of these substances act as beta-secretase inhibitors, inducers of IDE expression, GSK3B inhibitors, and mTOR inhibitors that can prevent AD progression in T2D patients. In addition, these compounds can act as NFkB inhibitors and stimulate Nrf2 activation to inhibit oxidative stress and neuroinflammation, which are major aspects of the pathogenesis of AD in T2D patients.

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References

Ahmad, B., Shah, M. & Choi, S. (2019). Oceans as a source of immunotherapy. *Marine Drugs*, **17**(5), 282. https://doi.org/10.3390/md17050282

Alonso, D., & Martnez, A. (2006). Marine Compounds as a New Source for Glycogen Synthase Kinase 3 Inhibitors. In A. Martinez, A. Castro. & M. Medina (Eds.), Wiley Series in Drug discovery and development (307–331). John Wiley & Sons, Inc. https://doi.org/10.1002/0470052171.ch16

Alzheimer's Association. (2022). 2022 Alzheimer's disease facts and figures. In *Alzheimer's and Dementia*, **18**(4). https://doi.org/10.1002/alz.12638

Ankisetty, S. Khan, S.I., Avula, B., Gochfeld, D., Khan, I.A., & Slattery, M. (2013). Chlorinated didemnins from the tunicate *Trididemnum solidum*. *Marine Drugs*, **11**(11), 4478–4486. https://doi.org/10.3390/md11114478

Arya, A., Nahar, L., Khan, H.U. & Sarker, S.D. (2020). Chapter Thirteen—Anti-obesity natural products. In S.D. Sarker & L. Nahar (Eds.), *Annual Reports in Medicinal Chemistry*, (55), 411–433. https://doi.org/10.1016/bs.armc.2020.02.006

Barbalace, M.C., Malaguti, M., Giusti, L., Lucacchini, A., Hrelia, S., & Angeloni, C. (2019). Anti-inflammatory activities of marine algae in Neurodegenerative Diseases. *International Journal of Molecular Sciences*, **20**(12), 3061. https://doi.org/10.3390/ijms20123061

Byun, E.B., Cho, E.J., Kim, Y.E., Kim, W.S., & Byun, E.H. (2018). Neuroprotective effect of polysaccharide separated from *Perilla frutescens* Britton var. Acuta Kudo against H₂O₂-induced oxidative stress in HT22 hippocampus cells. *Bioscience, Biotechnology, and Biochemistry,* **82**(8), 1344–1358. https://doi.org/10.1080/09168451.2018.1460572

Cai, Z., Chen, G., He, W., Xiao, M., & Yan, L.J. (2015). Activation of mTOR: A culprit of Alzheimer's disease? *Neuropsychiatric Disease and Treatment*, **11**, 1015–1030 https://doi.org/10.2147/NDT.S75717

Cardoso, M.J., Costa, R.R., & Mano, J.F. (2016). Marine Origin polysaccharides in drug delivery systems. *Marine Drugs*, **14**(2), 34. https://doi.org/10.3390/md14020034

Ghiciuc, C.M., Vicovan, A.G., Stafie, C.S., Antoniu, S.A, & Postolache, P. (2021). Marine-derived compounds for the potential treatment of glucocorticoid resistance in severe asthma. *Marine Drugs*, **19**(11), 586. https://doi.org/10.3390/md19110586

Gogineni, V., & Hamann, M.T. (2018). Marine natural product peptides with therapeutic potential: Chemistry, biosynthesis, and pharmacology. *Biochimica et Biophysica Acta (BBA) - General Subjects*, **1862**(1), 81–196. https://doi.org/10.1016/j.bbagen.2017.08.014

Granic, I. Dolga, A.M., Nijholt, I.M., Van Dijk, G., & Eisel, U.L.M. (2009). Inflammation and NF-κB in Alzheimer's

disease and diabetes. *Journal of Alzheimer's Disease*, **16**(4), 809–821. https://doi.org/10.3233/JAD-2009-0976

Guo, M., Zuo, L., Qiao, G., Liu, M., Cao, S., & Lin, X. (2021). PI3K/Akt/mTOR signaling as targets for developing anticancer agents from marine organisms. *Journal of Ocean University of China*, **20**(3), 688–694. https://doi.org/10.1007/s11802-021-4636-0

Hafez G.S., & Kijjoa, A. (2021). Marine-derived compounds with anti-Alzheimer's disease activities. *Marine Drugs*, **19**(8), 410. https://doi.org/10.3390/md19080410

He, L., & Sun, Y. (2021). The potential role of Keap1-Nrf2 pathway in the pathogenesis of Alzheimer's disease, type 2 diabetes, and type 2 diabetes-related Alzheimer's disease. *Metabolic Brain Disease*, **36**(7), 1469–1479. https://doi.org/10.1007/s11011-021-00762-z

Kamata, K., Okamoto, S., Oka, S., Kamata, H., Yagisawa, H., & Hirata, H. (2001). Cycloprodigiosin hydrocloride suppresses tumor necrosis factor (TNF) α -induced transcriptional activation by NF-κB. *FEBS Letters*, **507**(1), 74–80. https://doi.org/10.1016/S0014-5793(01)02946-5

Kim, S.Y., Ahn, G., Kim, H.S., Je, J.G., Kim, K.N., & Jeon, Y.J. (2020). Diphlorethohydroxycarmalol (DPHC) isolated from the brown alga ishige okamurae acts on inflammatory myopathy as an inhibitory agent of TNF-α. *Marine Drugs*, **18**(11), 529. https://doi.org/10.3390/md18110529

Kubota, T., Kurimoto, S., & Kobayashi, J. (2020). Chapter one—The manzamine alkaloids. In H.J. Knölker (Ed.), *The alkaloids: Chemistry and biology* (Vol. **84**(1–124). Academic Press. https://doi.org/10.1016/bs.alkal.2020.03.001

Lee, J., & Jun, M. (2019). Dual BACE1 and cholinesterase Inhibitory effects of phlorotannins from *Ecklonia cava*—An *in vitro* and *in silico* study. *Marine Drugs*, **17**(2). https://doi.org/10.3390/md17020091

Lei, M., Xu, H., Li, Z., Wang, Z., O'Malley, T.T., Zhang, D., Walsh, D.M., Xu, P., Selkoe, D.J., & Li, S. (2016). Soluble Aβ oligomers impair hippocampal LTP by disrupting glutamatergic/GABAergic balance. *Neurobiology of Disease*, **85**, 111–121.1 https://doi.org/10.1016/j.nbd.2015.10.019

Li, H.H., Su, J.H., Chiu, C.C., Lin, J.J., Yang, Z.Y., Hwang, W.I., Chen, Y.K., Lo, Y.H., & Wu, Y.J. (2013). Proteomic investigation of the sinulariolide-treated melanoma cells A375: Effects on the cell apoptosis through mitochondrial-related pathway and activation of caspase cascade. *Marine Drugs*, **11**(7), 2625–2642. https://doi.org/10.3390/md11072625

Li, Y.X., Himaya, S.W.A., & Kim, S.K. (2013). Triterpenoids of marine origin as anti-cancer agents. *Molecules*, **18**(7), 7886–7909. https://doi.org/10.3390/molecules18077886

Matos, A.M. de, Macedo, M. P. de, & Rauter, A.P. (2017). Bridging type 2 diabetes and Alzheimer's disease: Assembling the puzzle pieces in the quest for the molecules with therapeutic and preventive potential. *Medicinal Research Reviews*, **00**(0), 1–64. https://doi.org/10.1002/med

Motohashi, K., Toda, T., Sue, M., Furihata, K., Shizuri, Y., Matsuo, Y., Kasai, H., Shin-ya, K., Takagi, M., Izumikawa, M., Horikawa, Y., & Seto, H. (2010). Isolation and structure

elucidation of tumescenamides A and B, two peptides produced by *Streptomyces tumescens* YM23-260. *The Journal of Antibiotics*, **63**(9), 549–552. https://doi.org/10.1038/ja.2010.73

Mycroft-West, C.J., Cooper, L.C., Devlin, A.J., Procter, P., Guimond, S.E., Guerrini, M., Fernig, D.G., Lima, M.A., Yates, E.A., & Skidmore, M.A. (2019). A glycosaminoglycan extract from Portunus pelagicus inhibits BACE1, the β secretase implicated in Alzheimer's disease. *Marine Drugs*, **17**(5). https://doi.org/10.3390/md17050293

Mycroft-West, C.J., Devlin, A.J., Cooper, L.C., Procter, P., Miller, G.J., Fernig, D.G., Guerrini, M., Guimond, S.E., Lima, M.A., Yates, E.A., & Skidmore, M.A. (2020). Inhibition of BACE1, the β -secretase implicated in Alzheimer's disease, by a chondroitin sulfate extract from *Sardina pilchardus*. *Neural Regeneration Research*, **15**(8), 1546–1553. https://doi.org/10.4103/1673-5374.274341

Najem, D., Bamji-Mirza, M., Yang, Z., & Zhang, W. (2016). A β -induced insulin resistance and the effects of insulin on the cholesterol synthesis pathway and A β secretion in neural cells. *Neuroscience Bulletin*, **32**(3), 227–238. https://doi.org/10.1007/s12264-016-0034-9

Naushad, M., Durairajan, S.S.K., Bera, A.K., Senapati, S., & Li, M. (2019). Natural compounds with anti-BACE1 activity as promising therapeutic drugs for treating Alzheimer's disease. *Planta Medica*, **85**(17), 1316–1325. https://doi.org/10.1055/a-1019-9819

Nguyen, L.S., Vautier, M., Allenbach, Y., Zahr, N., Benveniste, O., Funck-Brentano, C., & Salem, J.E. (2019). Sirolimus and mTOR inhibitors: A review of side effects and specific management in solid organ transplantation. *Drug Safety*. https://doi.org/10.1007/s40264-019-00810-9

Nguyen, T.T., Ta, Q.T.H., Nguyen, T.K.O., Nguyen, T.T.D., & Giau, V. V. (2020). Type 3 diabetes and its role implications in Alzheimer's disease. *International Journal of Molecular Sciences*, **21**(9), 1–16. https://doi.org/10.3390/ijms21093165

Sharma, S., Guru, S.K., Manda, S., Kumar, A., Mintoo, M. J., Prasad, V.D., Sharma, P.R., Mondhe, D.M., Bharate, S.B., & Bhushan, S. (2017). A marine sponge alkaloid derivative 4-chloro fascaplysin inhibits tumor growth and VEGF mediated angiogenesis by disrupting PI3K/Akt/mTOR signaling

cascade. *Chemico-Biological Interactions*, **275**, 47–60. https://doi.org/10.1016/j.cbi.2017.07.017

Shieh, J.C.C., Huang, P.T., & Lin, Y.F. (2020). Alzheimer's Disease and diabetes: Insulin signaling as the bridge linking two pathologies. *Molecular Neurobiology*, *57*(4), 1966–1977. https://doi.org/10.1007/s12035-019-01858-5

Song, B., & Zhou, W. (2022). Amarogentin has protective effects against sepsis-induced brain injury via modulating the AMPK/SIRT1/NF-κB pathway. *Brain Research Bulletin*, **189**, 44–56.

https://doi.org/10.1016/j.brainresbull.2022.08.018

Soriano, S., Lu, D.C., Chandra, S., Pietrzik, C.U., & Koo, E.H. (2001). The amyloidogenic pathway of amyloid precursor protein (APP) is independent of its cleavage by caspases. *Journal of Biological Chemistry*, **276**(31), 29045–29050. https://doi.org/10.1074/jbc.M102456200

Tumminia, A., Vinciguerra, F., Parisi, M., & Frittitta, L. (2018). Type 2 diabetes mellitus and Alzheimer's disease: Role of insulin signalling and therapeutic implications. *International Journal of Molecular Sciences*, **19**(11). https://doi.org/10.3390/ijms19113306

Varga, E., Juhász, G., Bozsó, Z., Penke, B., Fülöp, L., & Szegedi, V. (2015). Amyloid-β1-42 disrupts synaptic plasticity by altering glutamate recycling at the synapse. *Journal of Alzheimer's Disease*, **45**(2), 449–456. https://doi.org/10.3233/JAD-142367

Wiese, J., Imhoff, J.F., Gulder, T.A.M. Labes, A., & Schmaljohann, R. (2016). Marine fungi as producers of benzocoumarins, a new class of inhibitors of glycogensynthase-kinase 3. Marine Drugs, **14**(11), Article 11. https://doi.org/10.3390/md14110200

Wrighten, S.A., Piroli, G.G., Grillo, C.A., & Reagan, L.P. (2009). A look inside the diabetic brain: Contributors to diabetes-induced brain aging. *Biochimica et Biophysica Acta - Molecular Basis of Disease*, **1792**(5), 444–453. https://doi.org/10.1016/j.bbadis.2008.10.013

Yu, J.H., Han, K., Park, S., Cho, H., Lee, D.Y., Kim, J.W., Seo, J.A., Kim, S.G., Baik, S.H., Park, Y.G., Choi, K.M., Kim, S.M., & Kim, N.H. (2020). Incidence and risk factors for dementia in type 2 diabetes mellitus: A nationwide population-based study in Korea. *Diabetes and Metabolism Journal*, **44**(1), 113–124. https://doi.org/10.4093/dmj.2018.0216